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(54) Title: USE OF A COL 1A1 MINI-GENE CONSTRUCT TO INHIBIT COLLAGEN SYNTHESIS (57) Abstract Methods are provided for controlling the synthesis of protein from the human gene for the pro α 1(I) chain of type I procollagen in cells using mini-gene constructs which are placed in modified viruses which are infected into cells which synthesize the protein. Methods for controlling collagen deposition in tissue and methods of treating conditions characterized by excessive deposition of collagen using the mini-gene constructs are also provided.		

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USE OF A COL1A1 MINI-GENE CONSTRUCT TO INHIBIT COLLAGEN SYNTHESIS

BACKGROUND OF THE INVENTION

A number of inherited diseases have recently been shown
5 to be caused by mutations in collagen genes that either cause
synthesis of insufficient amounts of collagen or defective
collagen, but other human diseases are caused by over-
production of normal collagen in a form referred to as either
fibrotic tissue or scars. Deposition of collagen is a normal
10 part of the repair process that occurs when any tissue is
injured. In man, however, the collagen deposition that occurs
is frequently excessive and irreversible. The result is
replacement of normal cells and tissues with inert collagen in
the form of tough fibers. There are many examples of this
15 excessive deposition of collagen. One example is injury to the
skin following trauma, burns or surgery. In normal people,
scars form because some of the cells in the skin (fibroblasts)
synthesize excessive amounts of collagen that are deposited as
scar tissue before the cells that synthesize the outer layers
20 of the skin (epithelial cells) can regenerate themselves and
form normal skin over the area of injury. In some individuals,
particularly Asians and Blacks, the deposition of collagen is
extreme so that large tumors consisting almost entirely of
collagen fibers called keloids and form at the site of even a
25 minor injury such as a small surgical incision and become a
source of serious disfigurement and discomfort. Excessive
deposition of collagen as scars of the skin is a major problem

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in some types of animal husbandry. Most horses, for example, form large collagen deposits called "proud flesh" with almost every injury below the knee and many have to be destroyed as a result. Excessive deposition of collagen in man also occurs

5 after injury or surgery of internal organs. For example, excessive fibrosis or scarring frequently occurs following injury or surgery of the hand and is a significant cause of permanent work disability among mechanics. Surgery to the back for herniated disks and related problems frequently fails to

10 improve the status of the patient because excessive deposits of collagen form around the sites of surgical repair and produce pain and dysfunction because they impinge on nerves. Excessive deposit of collagen fibers is also a major feature of cirrhosis of the liver. When the liver is damaged by viruses, alcohol or

15 toxic materials, a competition occurs between the replication of normal liver cells (hepatocytes) and collagen-producing cells that deposit collagen fibers. The deposition of collagen usually wins out in this competition and, as a result, the liver becomes non-functional and cirrhotic ("hard") because the

20 normal tissue is largely replaced by collagen fibers. A similar sequence of events is frequently seen in the lung where injury from a number of agents, including many agents used in cancer chemotherapy, is followed by replacement of normal lung tissue so that the lung becomes stiff and can no longer

25 exchange oxygen and other gases. The resulting condition (pulmonary interstitial fibrosis) is frequently lethal. A similar type of pulmonary fibrosis also occurs in some individuals for unknown reasons. In addition, excessive deposition of collagen fibers can be a general phenomenon of

30 some diseases. In the condition known as progressive systemic sclerosis, excessive amounts of collagen are deposited in the skin and many other organs. The patients are described as becoming "encased in leather". The result can be a total inability to use the hands, to open the mouth to eat, or to

35 absorb nutrients from the gastrointestinal tract. Many of the patients die because internal organs such as the kidney are replaced by collagens.

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Excessive deposition of collagen has also been implicated, but not demonstrated as directly, as a major debilitating event in atherosclerosis and arteriosclerosis where the "hardening" of the blood vessels is explained in part by the fact that they contain more collagen. Excessive deposition of collagen has also been implicated as a secondary explanation for the muscle failure in muscular dystrophy. Also, it has been implicated in some blood diseases in which the blood-producing cells of bone marrow are replaced by collagen fibers (myelofibrosis).

Overall, therefore, there are important needs for developing means of limiting and controlling the deposition of normal collagen in tissues. No effective measures are currently available.

15 SUMMARY OF THE INVENTION

A method for controlling the synthesis of protein from the human gene for the pro α 1(I) chain of type I procollagen in cells is provided comprising a gene selected from the group consisting of a mini-gene construct of the human gene for the pro α 1(I) chain of type I procollagen and a mini-gene construct of the human gene for the pro α 1(I) chain of type I procollagen with the second half of the gene sequence being antisense; placing one or both genes in a modified virus selected from the group consisting of a retrovirus and an adenovirus; and infecting cells capable of synthesizing protein from the human gene for the pro α 1(I) chain of type I procollagen with the modified virus containing the mini-gene construct or constructs.

The mini-gene contains an internal deletion of the COL1A1 gene whereby 42 exons (exons 4 to 46) and most of the associated introns have been removed.

A method for controlling collagen deposition in tissue is also provided comprising providing a gene selected from the group consisting of a mini-gene construct of the human gene for the pro α 1(I) chain of type I procollagen and a mini-gene construct of the human gene for the pro α 1(I) chain of type I

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procollagen with the second half of the gene sequence being antisense ; combining one or both mini-gene constructs with a selected delivery means; and delivering one or both mini-gene constructs to the tissue wherein the collagen deposition is to be controlled. In one embodiment, the delivery means comprises a composition containing a protein or lipid or other molecules chemically linked to one or both mini-gene constructs. In other embodiments, the delivery means comprises an electrical current, laser beam, or pressure. The mini-gene construct or constructs may be administered by injection, direct topical application, inhalation, or orally.

In another embodiment of the invention, a method of treating a mammal having a condition characterized by the excessive deposition of collagen in tissue comprising administering a therapeutically effective amount of a composition consisting essentially of a gene selected from the group consisting of a mini-gene construct of the human gene for the $\alpha 1(I)$ chain of type I procollagen and a mini-gene construct of the human gene for the $\alpha 2(I)$ chain of type I procollagen with the second half of the gene sequence being antisense and a pharmaceutically acceptable carrier to a mammal having a condition characterized by the excessive deposition of collagen in its tissue. The condition characterized by the excessive deposition of collagen in tissue may be a cirrhotic or fibrotic disease. Cirrhosis of the liver, pulmonary interstitial fibrosis, progressive systemic sclerosis, atherosclerosis, arteriosclerosis, and myelofibrosis may be treated with the methods of the invention. The invention may also be used to treat conditions resulting from injury to the skin of the mammal following trauma, burns, surgery or the result of injury or surgery of internal organs.

Composition of the invention may be administered by injection, direct application, inhalation, or orally.

DESCRIPTION OF THE DRAWINGS

Figure 1 shows the structure of the mini-gene construct. Figure 1(A) shows the structure of the mini-gene for the $\alpha 1(I)$

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1(I) chain of human type I procollagen. The gene contains 2,300 bp of the promoter region, about 2,000 bp of 3'-sequences, and all of the gene except for exons 6 to 46 and most of the associated introns. Because the deletion of coding sequences was in-frame, the deleted gene codes for a shortened 5 sequences was in-frame, the deleted gene codes for a shortened pro α 1(I) chain. Figure 1B shows the structure of the antisense mini-gene construct. The 3'-half of the mini-gene (A) has been inverted so that the second half of the gene has a base sequence that is complementary to the sequences for exons 47 to 10 52 and the associated introns of the human gene of pro α 1(I) chains of type I procollagen.

Figures 2A-D show typical pedigrees from experiments in which a line of transgenic mice expressing high levels of the mine-gene was bred with a line expressing the antisense gene. 15 As indicated, all mice inheriting the mini-gene only were dead at birth. All mice inheriting the antisense gene only were viable and had no phenotype. All mice inheriting both genes were also viable and without a phenotype.

Figure 3 shows the results of western blotting 20 experiments to demonstrate the effect of the antisense gene or expression of the mini-gene. Protein was extracted from samples of tail from mice breeding experiments such as those shown in Figure 2. The proteins were separated by polyacrylamide gel electrophoresis in sodium dodecyl sulfate, 25 the proteins were electroeluted onto filters, and the proteins were immunoblotted with a polyclonal antibody (rabbit) that reacts with the COOH-terminal propeptide in the pro α 1(I) chains of both mouse and human type I procollagen. Upper panel: Immunoblot from selected members of one litter. The bands (top 30 to bottom) in the immunoblot are pro α 1(I) chains of mouse type I procollagen; partially processed bands of mouse pro α 1(I) chains that retain the COOH-terminal propeptide; shortened pro α 1(I) chains of human type I procollagen [mini-pro α 1(I)]; and a non-specific band of protein detected by the antibody. 35 The results show that in the mouse inheriting both genes, there was a decrease in the steady-state level of intracellular shortened human pro α 1(I) chains [mini-pro α 1(I)] and an increase

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in mouse $\text{pro}\alpha 1(\text{I})$ and $\text{pCa1}(\text{I})$ chains. Lower panel: Similar results from a s cond litter.

DETAILED DESCRIPTION OF THE INVENTION

The present invention offers methods for controlling
5 collagen deposition in tissues. Mini-gene constructs useful in
the methods of the invention are shown in Figure 1. The mini-
gene construct (Figure 1) contains the first five exons and the
last six exons of the gene together with 2.3 kb of the 5'-
flanking sequence and about 2 kb of the 3'-flanking sequence.
10 The 5' and 3' fragments of the gene are joined through an
artificial SalI site located within two intervening sequences
so as not to alter any sequences required for RNA splicing.
Also, all the coding sequences were in frame. The structure of
the construct was verified by mapping with restriction
15 endonucleases and partial DNA sequencing. The 5'-end of the
gene has an artificial NotI site. Thus, the construct can be
excised cleanly from the plasmid vector with a NotI/BamHI
restriction enzyme digestion.

Figure 1A shows a mini-gene construct of the human gene
20 for the $\text{pro}\alpha 1(\text{I})$ chain of type I procollagen (COL1A1). The
gene is denoted a "mini-gene" because it contains an internal
deletion whereby 42 exons (exons numbers 6 to 46) and most of
the associated introns have been removed from the gene. This
mini-gene has been used to produce transgenic mice that can be
25 used for testing therapies for the treatment of osteogenesis
imperfecta and osteoporosis, as described in Applicants'
copending application Serial No. 713,945. It was designed to
cause synthesis of shortened $\text{pro}\alpha 1(\text{I})$ chains of type I
procollagen that bind to normal chains of the same protein and
30 cause the degradation of the normal chains because a molecule
containing both shortened $\text{pro}\alpha 1(\text{I})$ chains, and normal $\text{pro}\alpha 1(\text{I})$
chains cannot fold into a biologically functional molecule. As
shown by Example 1 herein, expression of the mini-gene in
transgenic mic produces a net decrease in the amount of type
35 I collagen found in tissues such as bone that are normally rich
in the protein. In th present invention, the sam mini-gene

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is used to prevent excessive deposition of collagen by placing it in a modified virus or other composition that will serve to carry the mini-gene to cells producing excessive amounts of collagen and cause synthesis of shortened pro α 1(I) chains in the cells. As a result, the amount of collagen synthesized and deposited as collagen fibers by the cells decreases.

Another embodiment is illustrated in Figure 1B. Here, the second half of the mini-gene is inverted so that the second half codes for complementary or antisense sequences of RNA bases for exons 47 to 52 and the associated introns. Because the second half of the "antisense gene" codes for a complementary sequence, the RNA transcribed from the gene will bind to the RNA transcribed from a normal gene for pro α 1(I) chains of the same cell. Because part of the normal RNA is rendered double stranded, it cannot be used to synthesize protein. Instead, it either becomes trapped in the nucleus of the cell or is degraded, probably by an enzyme that specifically degrades double-stranded DNA such as ribonuclease H. As shown by Example 2, expression of the antisense gene in transgenic mice can inhibit expression of a human gene for the same protein in a highly specific manner. Therefore, the method for inhibiting excessive deposit of collagen is to place the antisense gene in a modified virus or other composition that will serve to carry the antisense gene to cells producing excessive amounts of collagen and cause synthesis of an antisense RNA in the cells and, therefore, reduce the amount of collagen synthesized and deposited as collagen fibers by the cells.

Similar means can be used to deliver for the mini-gene and antisense gene to cells. For example, either one or both of the genes can be placed in modified RNA virus (retrovirus) that is engineered so that the virus will infect cells but will not generate any replicative virus that can infect a second generation of cells. Specific examples of the design and use of such retroviruses for the delivery of genes to cells are described in Tolstoshev, P., *Bone Marrow Transplantation* 1992, 9, 148-150 and Morgan, R.A. et al., *Nucleic Acids Research*

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1992, 20, 1293-1299. As another example, either on or both of
th genes can be placed in an appropriately modified virus that
in its native form causes respiratory diseases (adenovirus) and
the modified virus containing either gene can be used to infect
5 cells producing excessive amounts of collagen. Examples of the
use of such adenoviruses to deliver genes to cells are
described in Engelhardt and Wilson, J. Pharmacy and
Pharmacology 1992, 44, 165-167; Yoshimura et al., Nucleic Acids
Research 1992, 20, 3233-3240 and Crystal, R. Am J of Medicine
10 1992, 92, 445-555.

The mini-gene and the antisense gene constructs are
specifically designed to control the synthesis of protein from
the human gene for the pro α 1(I) chain of type I procollagen.
Controlling the synthesis of protein from this gene is
15 appropriate because the vast majority of the collagen found in
scars and fibrotic tissues is type I collagen which cannot be
synthesized by cells without pro α 1(I) chains of the precursor
molecule known as type I procollagen. The present invention,
however, also includes other versions of mini-gene constructs
20 in which other regions are deleted or specific base sequences
are replaced. In particular, it may be useful to replace the
promoter region (from exon 1 to -2.3 kb) with other promoters
or combinations of promoters and enhancers from other genes
that will improve transport to, or the controlled expression
25 of, the gene in specific cells. Similar variations of the
design of the antisense gene are also contemplated. Also, in
addition to viruses as vectors for delivery of the genes, the
present invention also contemplates other means of delivery
such as incorporation of one or both genes into compositions
30 that will facilitate entry of the genes and facilitate control
of their expression in cells. For example, it may be useful to
chemically link one or both kinds of genes to proteins, lipids
or other molecules that will facilitate binding of the genes to
specific cells and entry into their nuclei. Also, it may be
35 us ful to encapsulate one or both kinds of genes in lipids or
viral capsules or similar molecules for the same purpose. The
present invention also includes delivery of one or both kinds

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of genes to cells by means assisted by physical techniques such as electrical currents, ionic beams, laser beams or pressure. In using both kinds of genes to control fibrosis, it is contemplated that they can be used both for systemic administration to man and other animals and for local administration. For example, one or both kinds of genes in an appropriate composition can be administered by injection either into a vein, a muscle or under the skin. In an appropriate composition such as a virus vector, they can be administered by mouth. Also, they can be administered by direct application to the site of an injury or surgical incision in tissues such as the skin, the hand, or the back, to limit the amount and extent of the fibrosis and scarring that will occur. In addition, they can be administered to the lungs by inhalation as aerosols. For direct application to tissues, the genes can be in compositions that are in solution, in particulate suspensions, in oily compounds, in gels, or in solid materials that either remain in the tissues for extended periods of time or are rapidly absorbed or degraded by the tissues.

The invention is further illustrated by the following, non-limiting examples.

EXAMPLES

Example 1

A series of experiments were carried out to demonstrate that the mini-gene for human pro α 1(I) chains (Figure 1A) can be used to specifically control the amount of type I collagen fibers deposited in tissues.

Lines of transgenic mice expressing the mini-gene were prepared by injecting fertilized eggs from matings of inbred FVB/N males and females, and the eggs were transferred either on the same day or cultured to the two cell stage and then transferred to the oviduct of a 0.5 day post-coitus pseudo-pregnant female. To identify progeny that contained the transgene in their genomes, DNA in a sample of tail or other tissue was analyzed for presence of the mini-gene as described in Examples 3 and 4.

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10

TABLE I
SUMMARY OF MOUSE PHENOTYPES AT BIRTH

15	Mice		Phenotype of transgenic mice ^a		
	Total	Transgenic	Lethal with fractures	Non-lethal with fractures	No fractures
idCOL1xWt ^b	195	89	6%	33% ^c	61% ^c
idCOL1xidCOL1	70	48	40%	ND ^d	ND ^d

^b Matings of transgenic males (idCOL1) with wild-type (wt) females from the same line of mice (FVB/N).

^c Values based on x-ray examination on day of birth of 15 viable transgenic mice from 5 litters.

^d Not determined.

To assay the collagen content of tissues from the mice, the femurs were selected as tissues that can be clearly dissected from the animals in a manner that is largely free of other tissues and the total collagen content of the tissue readily compared among groups of animals. As indicated In Table II, the collagen content of the femurs in the transgenic mice was significantly less than the collagen content of femurs from control littermates that did not have the mini-gene in their genomes. The results directly

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demonstrated that the mini-gene can be used to limit the amount of normal collagen deposited in tissues in mammals.

TABLE II

COMPOSITION OF FEMURS FROM 6-WEEK OLD MALE MICE^a

	Length (mm)	Wet weight (mg)	Mineral (mg)	Collagen (mg)
5 Controls (n=22)	14.1±0.39	58.46±7.39	19.9±1.87	4.3±1.16
10 Transgenics (n=16)	12.4±1.38**	42.4±2.12**	13.6±5.39**	2.9±1.24**

^a Values are means ± standard deviation.

** p<0.001 by the Student t-test.

Example 2

15 An experiment was carried out to demonstrate that the antisense gene (Figure 1B) can be used to limit and control the synthesis of pro α 1(I) chains of type I procollagen. The design of the experiment was based on the availability of two different lines of transgenic mice, one line expressing high
20 levels of a human mini-gene for the pro α 1(I) chains of type I procollagen (Figure 1A) and the other line expressing an antisense version of the same gene (Figure 1B). The line expressing high levels of the human mini-gene developed a lethal phenotype in that about 90% of the mice died at birth,
25 because the mini-gene caused synthesis of shortened pro α 1(I) chains that caused degradation of the normal pro α 1(I) and pro α 2(I) chains synthesized from the endogenous mouse genes. As a result, there was a marked decrease in the amount of type I collagen present in their tissues and their bones and other
30 collagen-rich tissues were extremely fragile. In contrast, the transgenic mice expressing the antisense version of the same gene (Figure 1B) were viable and showed no phenotype such as increased fragility of bones and other tissues. The lack of a phenotype in the transgenic mice was explained by the fact that

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the antisense gene, like the mini-gene, was a human gene and the human and mouse genes have a number of differences in their base sequences. Because of the differences, the RNA from the antisense gene does not significantly bind to the RNA from the mouse gene for pro α 1(I) chains and, therefore, does not reduce synthesis of type I collagen from the endogenous mouse genes. The experiment here consisted of breeding the small number of viable transgenic mice expressing high levels of the human mini-gene with transgenic mice expressing the antisense version of the same human gene. The question addressed in the experiment was whether mice that inherit both genes show decreased synthesis of shortened pro α 1(I) chains from the human mini-gene.

Transgenic mice from the two lines were bred and the progeny analyzed both for the presence of the two genes in their genomes and the synthesis of pro α 1(I) chains in their tissues. The presence of the genes was detected by Southern blot analysis of DNA from tail or other tissues with 32 P-labeled DNA probes for specific fragments of the genes. The synthesis of pro α 1(I) chains was assayed by the steady-state intracellular levels of the chains as detected by Western blotting of protein extracts from tissues with polyclonal antibodies from rabbits that reacted with the COOH-terminal propeptides of pro α 1(I) chains from both the mouse gene and the human mini-gene.

A total of ten litters and 77 mice were obtained. Thirty-one of the mice had a normal genotype, 22 inherited the antisense gene only, 13 inherited the mini-gene only, and 11 inherited both genes. Twelve of the 13 mice inheriting the mini-gene only had the lethal phenotype. In contrast, only 3 of 11 of mice with both genes had a lethal phenotype.

As indicated by the typical pedigree(s) in Figure 2A-D, most of the mice that inherited only the mini-gene died at birth. In contrast, most of the mice that inherited both genes survived. As indicated by the examples in Figure 3, there was a marked decrease in the synthesis of shortened human pro α 1(I) chains in tissues from mice that inherited both genes.

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Moreover, there was an increase in the normal pro α 1(I) chains synthesized from the endogenous mouse gene. Therefore, the results demonstrated that expression of the human antisense gene decreased synthesis of shortened pro α 1(I) chains from the human mini-gene in a highly specific manner.

X-ray examination of the mice revealed that all the mice with the mini-gene and the lethal phenotype had extensive fractures of long bones and ribs. Of the 11 mice with both genes, the two that died on the day of birth also had extensive fractures of long bones and ribs. The fractures in a third mouse that died on day 2 could not be evaluated because of maternal cannibalism. Therefore, the value of three out of 11 or 27% for a lethal phenotype in mice with both genes may be a slight overestimate. Three of the 11 mice with both genes had fractures of ribs that healed with callus formation when examined at one week. The remaining five of the 11 mice with both genes had no evidence of fractures.

To assay the effects of the antisense gene on expression of the mini-gene, extracts of tail from the mice were assayed by Western blotting with an antibody that cross-reacted with the full-length pro α 1(I) chains synthesized from the human mini-gene. In 11 mice that inherited the mini-gene only and developed the lethal phenotype, there was an apparent decrease in the mouse pro α 1(I) chains as indicated by a comparison of lanes in which about the same amounts of protein were loaded. More quantitative estimates were obtained by comparison of the ratio of mouse pro α 1(I) chains to mini-pro α 1(I) chains within the same sample and under conditions in which assays of the ratio are linear. The ratio was 0.64 ± 0.128 S.D. (n=11) in mice within the mini-gene and the lethal phenotype. In contrast, the ratio was 1.23 ± 0.686 S.D. (n=6) in mice with both genes and the normal phenotype. The difference in the mean was significant with a p value of 0.019 as estimated by the cumulative distribution of the standard normal distribution and a one-tail test. Tissue samples adequate for assay were available only on one of the three mice that inherited both genes and that had the lethal phenotype. In this one mouse,

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the ratio of normal to mini-pro α 1(I) chains was in the lower rang of the values for transgenic mice that inherited the mini-gene only and had a lethal phenotype with extensive fractures.

- 5 As indicated in Table III, the effect of the human antisense gene was consistently observed in a series of breeding experiments in that there was a consistent decrease in the ratio of shortened pro α 1(I) chains to normal pro α 1(I) chains in mice that inherited both genes. Also, as expected,
10 there was a consistent decrease in the lethal phenotype.

TABLE III
Effect of Antisense Gene on Lethal Phenotype

<u>Genotypes</u>					<u>Phenotype (%)</u>	
<u>Matings</u>	<u>Normal</u>	<u>Mini-Gene</u>	<u>Anti-sense gene</u>	<u>Both trans-genes</u>	<u>Normal</u>	<u>Lethal</u>
15 Mini-gene	44				100	
X wild type		30			13	87
Mini-gene	31				100	
20 X antisense			22		100	
gene		13			8	92
				11	73	27 ^a

- ^a Two mice died in a few hours and had extensive fractures of
25 ribs and long bones. A third mouse died on the second day. It was smaller than littermates but because of maternal cannibalism, the phenotype of bony fractures could not be evaluated fully. However, it is listed here as a lethal phenotype.

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Tabl IV

Phenotype of Transgenic Mice with B th the Mini-Gene and the Antisense Gene.

5	Number of mice <u>Normal</u>	<u>Genotype</u>		Ratio normal pro α 1 to mini- pro α 1 ^a	<u>Phenotype^b</u>		
		<u>Mini- gene</u>	<u>Antisense gene</u>		<u>Lethal w/ extensive fractures</u>	<u>Viable with fractures</u>	
10	10 7	+	+	0.64 \pm 0.128 1.23 \pm 0.686	10 1 ^c	3	3

^a Assayed by laser densitometry of Western blots similar to those shown in Figure 5.

15 ^b Fractures in viable mice were assessed by x-ray analysis within the first week of life.

^c Ratio of normal pro α 1(I) to mini-pro α 1(I) chains in mouse with the lethal phenotype was <0.39.

Overall, the results demonstrate that the human antisense
20 gene can be used as a highly specific means of limiting and
controlling synthesis by cells of the pro α 1(I) chain of type I
procollagen that is essential to form the type I collagen and
that is the major constituent of fibrous tissues and scars. Of
special importance here is the observation that expression of
25 the antisense gene did not significantly decrease synthesis of
normal pro α 1(I) chains from the mouse gene. This result
suggests that this action of the antisense gene is highly
specific and that its expression is unlikely to have much
effect on the expression of most normal genes whose base
30 sequences are far more different from those of the human
pro α 1(I) chains.

Example 3 Assays of Expression in Transfected Cells and Transgenic Mice

Assays of DNA and RNA

35 To assay for the presence of exogenous genes, standard Southern blotting is used (Maniatis et al., *Molecular Cloning*,

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A Laboratory Manual 1982). Restriction enzymes that give different size fragments from the exogenous and endogenous gene are identified. A sample of tail or other tissue was minced in 0.1 M NaCl, 0.1 M EDTA and 1% SDS in 50 mM Tris-HCl buffer (pH 7.5) containing 0.7 mg/ml proteinase K and incubated with agitation at 55°C for 8 h to 15 h. The digested sample was extracted with phenol, phenol/chloroform/isoamyl alcohol and then with chloroform/isoamyl alcohol (24:1). The DNA was ethanol precipitated and dissolved in 10 mM Tris-HCl buffer (pH 7.5) and 1 mM EDTA. The isolated DNA was digested either with EcoRI, KpnI/Clal, or HindIII. The digested samples were electrophoresed on 0.8% agarose gel and transferred onto a nitrocellulose filter. Filters were probed with the ³²P-labeled mini-gene. Copy number of the type I mini-gene in transgenic mice was assayed by simultaneously probing the filters of HindIII digested DNA with the mini-gene or a cDNA (Hf677) for the human pro α 1(I) chain. The two DNA probes were mixed in equal amounts before labeling by nick translation with ³²P to ensure about the same specific activity. X-ray films of the Southern blots were scanned on a densitometer to estimate the ratio of the exogenous and endogenous genes. For assay of gene copy number for the human pro α 1(II) mini-gene the filters were probed simultaneously with a 10 kb EcoRI-EcoRI fragment from the human gene and a 15 kb EcoRI-EcoRI fragment of the mouse gene for type II procollagen.

To assay expression of mini-genes as mRNA, standard Northern blot procedures are used. For RNA assays, total cellular RNA was isolated from tissues using the standard guanidine thiocyanate/cesium chloride procedure. Samples containing 3 to 10 μ g of RNA were electrophoresed on 1% agarose formaldehyde gels and transferred to nitrocellulose filters. To detect expression of human exogenous genes as mRNAs, probe-protection experiments or a single probe that has only selective regions of identity with mouse or human mRNAs are employed. For example, the filters were hybridized with 1.5 kb insert from Hf677, a cloned cDNA for the human pro α 1(I) chain. Alternatively, an EcoRI/PvuII fragment of 563 nucleotides from

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the 5'-end of a human cDNA for the pro α 1(I) chain has been used in S1-nuclease probe-protection assays. The probe is fully protected by human mRNA for type I procollagen, but generates a distinctive fragment of 153 nucleotides with mouse mRNA. The probe is prepared as a single-stranded anti-sense RNA by subcloning into M13. The probe is labeled by using universal primer, the product is cleaved with restriction endonuclease, and the single-stranded probe isolated by polyacrylamide gel electrophoresis.

10 Assay of Protein Expression

Assays for expression of the genes at the protein level may also be performed. DNA transfected fibroblasts are taken directly for these assays. For transgenic mice, tissues were assayed directly or cultured fibroblasts or chondrocytes were prepared. For assay in mouse tissues about 50 mg of tissue was homogenized in 0.5 ml of 4 M guanidine thiocyanate, 0.25 M EDTA, 10 mM N-ethylmaleimide and 1 mM p-aminobenzamidine. For examination of reduced proteins, the homogenizing buffer also contained 10 mM 2-mercaptoethanol, a dithiothreitol (DDT). For examination of unreduced proteins, the buffer contained 250 mM iodoacetamide in place of the 2-mercaptoethanol or dithiothreitol (DDT). The sample was homogenized with teflon glass homogenizer and clarified by centrifugation at 13,000 x g for 10 minutes. The sample was dialyzed against 0.15 M NaCl, 10 mM EDTA, 1 mM N-ethylmaleimide, and 0.3 mM p-aminobenzamidine in 50 mM Tris-HCl buffer (pH 7.4). Aliquots of 15 μ l were mixed with 15 μ l glycerol and 3 μ l 1% SDS and 0.0015% bromphenol blue containing either 5% 2-mercaptoethanol or 250 mM iodoacetamide. The samples were heated at 100°C for 5 minutes and separated by electrophoresis on 10% polyacrylamide gel in a mini-gel apparatus (BIORAD, Prot in II). The protein was electroeluted onto nitrocellulose filters and the filters were reacted with polyclonal rabbit antibodies specific for the human pro α 1(I) chain, specific for the human and mouse pro α 1(I) chain, or specific for the human pro α 1(II) chain. The secondary antibodies were anti-rabbit IgG coupled to alkaline phosphatase.

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C 11 Culturing and Labeling of Newly Synthesized Proteins

Skin fibroblasts from transgenic mice and control mice were grown at passage 5 in 25 cm² plastic flasks (Falcon Labware) in Dulbecco's modified Eagle's medium with 10% fetal calf serum. At confluency, the cells were incubated for 4 hours in 1.5 ml of fresh medium containing 50 µg/ml ascorbate and 10 µCi/ml ¹⁴C-proline (250 mCi/mmol, Amersham Corp.). At the end of the labeling period, 0.15 ml of solution containing 250 mM EDTA, 100 mM ethylmaleimide, 10 mM phenylmethanesulfonyl fluoride and 0.1% NaN₃ was added to each flask to prevent proteolysis. Immediately thereafter, the media was collected and the cell layer was washed three times with 2 ml of cold phosphate-buffered saline. Media was centrifuged for 5 minutes at 13,000 x g at 4°C. Proteins were precipitated by two volumes of cold ethanol for 1 hour at -20°C. The precipitate was collected by centrifugation at 13,000 x g for 30 minutes at 4°C, washed twice with 70% cold ethanol, and vacuum dried.

Cells were recovered from the flasks by adding 2 ml of a solution containing 50 mM ethylmaleimide, 1 mM phenylmethanesulfonyl fluoride and 0.01% NaN₃. The samples were incubated for 5 minutes at 4°C, frozen at -20°C, defrosted, scraped, transferred to a tube and vortexed. Insoluble material from lysate was removed by brief centrifugation. The supernatant was precipitated with two volumes of ethanol as described for media proteins.

Newly synthesized protein chains are analyzed by SDS-polyacrylamide gel electrophoresis before and after a series of protease digestions. For direct assay of the protein synthesized, the medium is removed, a cocktail of protease inhibitors added, and the protein precipitated with ammonium sulfate. The precipitated proteins are analyzed by SDS-polyacrylamide gel electrophoresis. Samples of cell layers are homogenized in buffer containing protease inhibitors and analyzed by SDS-polyacrylamide electrophoresis. For analysis of the proteins from transgenic mice containing the proα1(I) mini-gene, one-dimensional electrophoresis was carried out with SDS in a 4 to 15% gradient polyacrylamide gel. Two-dimensional

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electrophoresis may also be carried out with unreduced samples separated in one direction and reduced samples in a second direction. For two-dimensional analysis, the unreduced media or cellular proteins were first run in 7% polyacrylamide gel. Individual lanes were cut out from the first dimensional gel, treated with 5% 2-mercaptoethanol for 1 hour, and separated by electrophoresis in second dimension in a 12% polyacrylamide gel. Western blot analysis with polyclonal antibodies was carried out as described above. After detection of bands using antibodies, the same blots were exposed on x-ray film to detect ¹⁴C-labeled proteins and verify the identity of the pro α chains. Fluorograms of exposed films are evaluated for size of polypeptide chain synthesis, post-translational overmodification, and relative amounts of polypeptide synthesized.

Pro α chains and procollagen synthesized by the fibroblasts are also evaluated by SDS-polyacrylamide electrophoresis after fragmentation by pepsin, vertebrate collagenase or cyanogen bromide. In the case of exogenous genes in which new codons for cysteine have been introduced, the presence of dimers will be detected by examining fragments of pro α chains with or without reduction prior to electrophoresis.

Assay of Exogenous Gene Products by

Immunoprecipitation or Western Blotting

The protein on the SDS-acrylamide gels was electroeluted onto a nitrocellulose filter. The filter was then reacted with either a polyclonal antibody specific for the N-propeptide of the human pro α 1(I) chain or a polyclonal antibody that reacted with the C-propeptide of the pro α 1(I) chain from both mouse and human. The secondary antibody was anti-rabbit IgG coupled to alkaline phosphatase (Promega Biotech). To compare the relative amounts of protein synthesized from the mini-gene and the endogenous pro α 1(I) gene, the Western blots prepared with cross-reacting antibodies were photocopied onto transparencies and the transparencies were assayed with a densitometer. The

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same filter was assayed with varying intensity settings for the photoc pier in order to establish that the film response was linear. The observed values for the peaks of mini-gene and endogenous gene products per mg tissue varied with the age of the mouse, apparently because of the increase in the extracellular matrix in most tissues with age. However, values for the ratio of mini-gene product to endogenous gene product was relatively constant for transgenic mice from the same line, and repeated assays of the same sample of tissue gave a standard deviation of $\pm 25\%$ of mean ($n = 5$).

**Example 4 Evaluation of Bones and Other Tissues
in Transgenic Mice**

Bone status in lines of transgenic mice is evaluated by determinations of ash weight, single photon absorptiometry, mechanical testing, staining and microradiography.

Ash Weight Determination

Bones to be determined for ash weight testing are cleaned mechanically of soft tissue and a wet weight obtained and volume obtained by displacement for significant dehydration of the bone. Dry weight and ash weights are obtained as follows: The bone is placed in a crucible and dried in a muffle furnace at 100°C until the bone weight changes by less than a percent between determination. The bone is then incinerated in the muffle oven at $580-600^{\circ}\text{C}$ for 24 hours and ash weights are obtained. Ash weight can then be described both as total ash weight and ash weight per volume (the ratio of the ash weight to the initial moist volume of the bone).

Single Photon Absorptiometry

Single photon absorptiometry is performed on a standard Lunar single photon absorptiometer with a scan speed of .25 mm per second. Whole bone determination and single scan line determination to specific positions along the bone are obtained from evaluation of these scans. Scans are performed by immersing the bone in water in a standard plastic tray. Polycarbonate plastic stops have been glued to the tray to allow reproducible positioning of the bone samples. Manual

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bone edge detection is done for final bone mass determination by single photon absorptiometry.

Skeletal Morphology and Histology

To examine the skeletal morphology of transgenic mice containing the pro α 1(I) mini-gene, F₁ mice from the R-line were stained with Alizarin red and Alizarin blue in accordance with methods known in the art.

To examine the skeletal morphology and histology of transgenic mice having the pro α 1(II) mini-gene construct mineralized tissue of selected newborn mice was stained with Alizarin Red S. For microscopy of tissues, whole embryos were perfused with 2% paraformaldehyde and 2.5% glutaraldehyde in 0.1 M phosphate buffer (pH 7.4) shortly after birth. Tissues were decalcified by incubation in 10% EDTA (pH 7.4) for 2 to 6 days. Samples were post-fixed in 1% osmium tetroxide in 0.15 M cacodylate buffer (pH 7.4) for 2 to 4 hours at 20°C, washed three times with the cacodylate buffer, and then placed in 1% tannic acid in 0.15 M cacodylate buffer (pH 7.4). For polarized light microscopy, specimens embedded in paraffin were stained with Dirius Red (picrosirius staining) after incubation in xylol at 37°C overnight and with 2 mg/ml hyaluronidase (Sigma, St. Louis) in 0.1 M phosphate buffer (pH 6.9) at 37°C overnight. The sections were analyzed with an Ortholux 2 Pol-Bk microscope (Ernst Leitz GmbH, Wetzlar, FRG) operated in monochromatic light [λ = 543 nm; filter IL 543 (Schott, Mainz, FRG)] and using the de Senarmont compensation technique. For electron microscopy, samples embedded in Epon were cut into 50 to 60 nm sections, stained with uranyl acetate for 30 minutes and lead citrate for 2 to 4 minutes, and examined with a JEM-1200 EX electron microscope (Jeol Ltd., Tokyo, Japan)

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What is Claim d is:

1. A method for controlling the synthesis of protein from the human gene for the pro α 1(I) chain of type I procollagen in cells comprising:
 - 5 providing a gene selected from the group consisting of a mini-gene construct of the human gene for the pro α 1(I) chain of type I procollagen and a mini-gene construct of the human gene for the pro α 1(I) chain of type I procollagen with the second half of the gene sequence being antisense;
 - 10 placing one or both genes in a modified virus selected from the group consisting of a retrovirus and an adenovirus; and
 - 15 infecting cells capable of synthesizing protein from the human gene for the pro α 1(I) chain of type I procollagen with the modified virus containing the mini-gene construct or constructs.
2. The method of claim 1 wherein said mini-gene construct does not contain exons 6 to 46 of the human COL1A1 gene.
- 20 3. The method of claim 1 wherein the antisense half of the mini-gene construct codes for an antisense sequence for exons 47 to 52 of the COL1A1 gene and the associated introns.
4. A method for controlling collagen deposition in tissue comprising:
 - 25 providing a gene selected from the group consisting of a mini-gene construct of the human gene for the pro α 1(I) chain of type I procollagen and a mini-gene construct of the human gene for the pro α 1(I) chain of type I procollagen with the second half of the gene sequence being antisense;
 - 30 combining one or both mini-gene constructs with a selected delivery means; and
 - delivering one or both mini-gene constructs to the tissue wherein the collagen deposition is to be controlled.

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5. The method of claim 4 wherein said mini-gene construct does not contain exons 6 to 46 of the human COL1A1 gene.

6. The method of claim 4 wherein the antisense half of the mini-gene construct codes for an antisense sequence for exons 47 to 52 of the human COL1A1 gene and the associated introns.

7. The method of claim 4 wherein said delivery means comprises a composition containing a protein or lipid chemically linked to one or both mini-gene constructs.

8. The method of claim 4 wherein said delivery means comprises an electrical current, ionic beam, laser beam, or pressure.

9. The method of claim 4 wherein said delivering step comprises administering the mini-gene construct or constructs by injection, direct topical application, inhalation, or orally.

10. A method of treating a mammal having a condition characterized by the excessive deposition of collagen in tissue comprising:

administering a therapeutically effective amount of a composition consisting essentially of a gene selected from the group consisting of a mini-gene construct of the human gene for the $\text{pro}\alpha 1(\text{I})$ chain of type I procollagen and a mini-gene construct of the human gene for the $\text{pro}\alpha 1(\text{I})$ chain of type I procollagen with the second half of the gene sequence being antisense and a pharmaceutically acceptable carrier to a mammal having a condition characterized by the excessive deposition of collagen in its tissue.

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11. The method of claim 10 wherein said mini-gene construct does not contain exons 6 to 46 of the human COL1A1 gene.
12. The method of claim 10 wherein the antisense half
5 of the mini-gene construct codes for an antisense sequence for exons 47 to 52 of the human COL1A1 gene and the associated introns.
13. The method of claim 10 wherein the condition
10 characterized by the excessive deposition of collagen in tissue is a cirrhotic or fibrotic disease.
14. The method of claim 10 wherein the condition is
selected from the group consisting of cirrhosis of the liver,
pulmonary interstitial fibrosis, progressive systemic
sclerosis, atherosclerosis, arteriosclerosis, and
15 myelofibrosis.
15. The method of claim 10 wherein the condition is the
result of injury to the skin of the mammal following trauma,
burns, or surgery.
16. The method of claim 10 wherein the condition is the
20 result of injury or surgery of internal organs of the mammal.
17. The method of claim 10 wherein the composition is
administered by injection, direct application, inhalation, or
orally.
18. The method of claim 10 wherein the mammal is a
25 human.

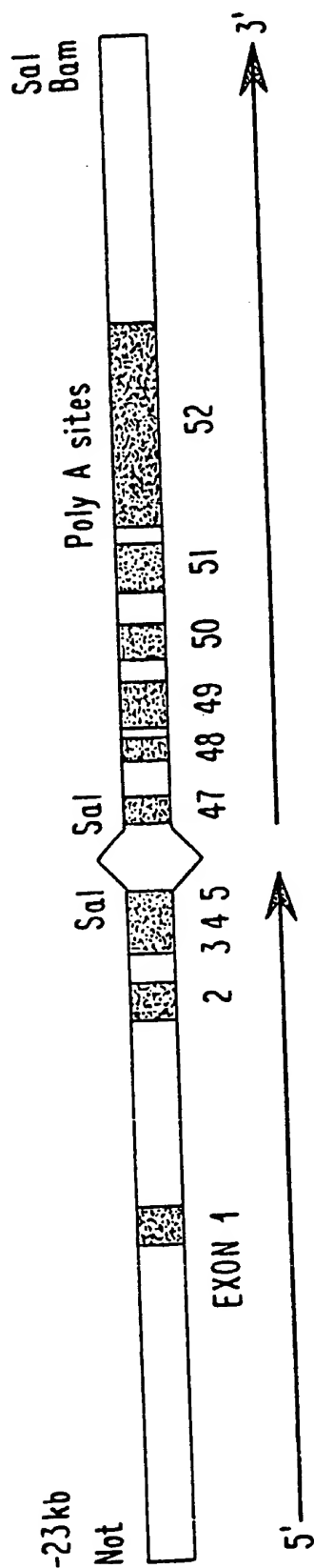


Fig. 1A

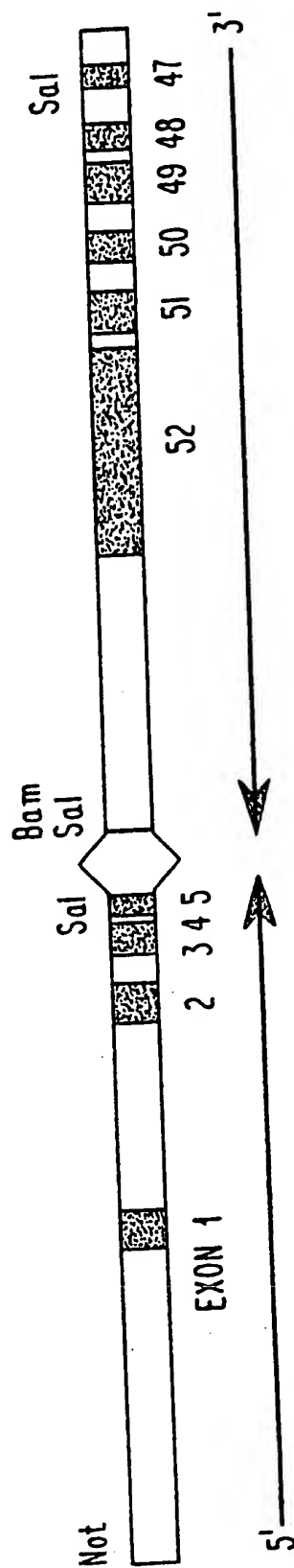
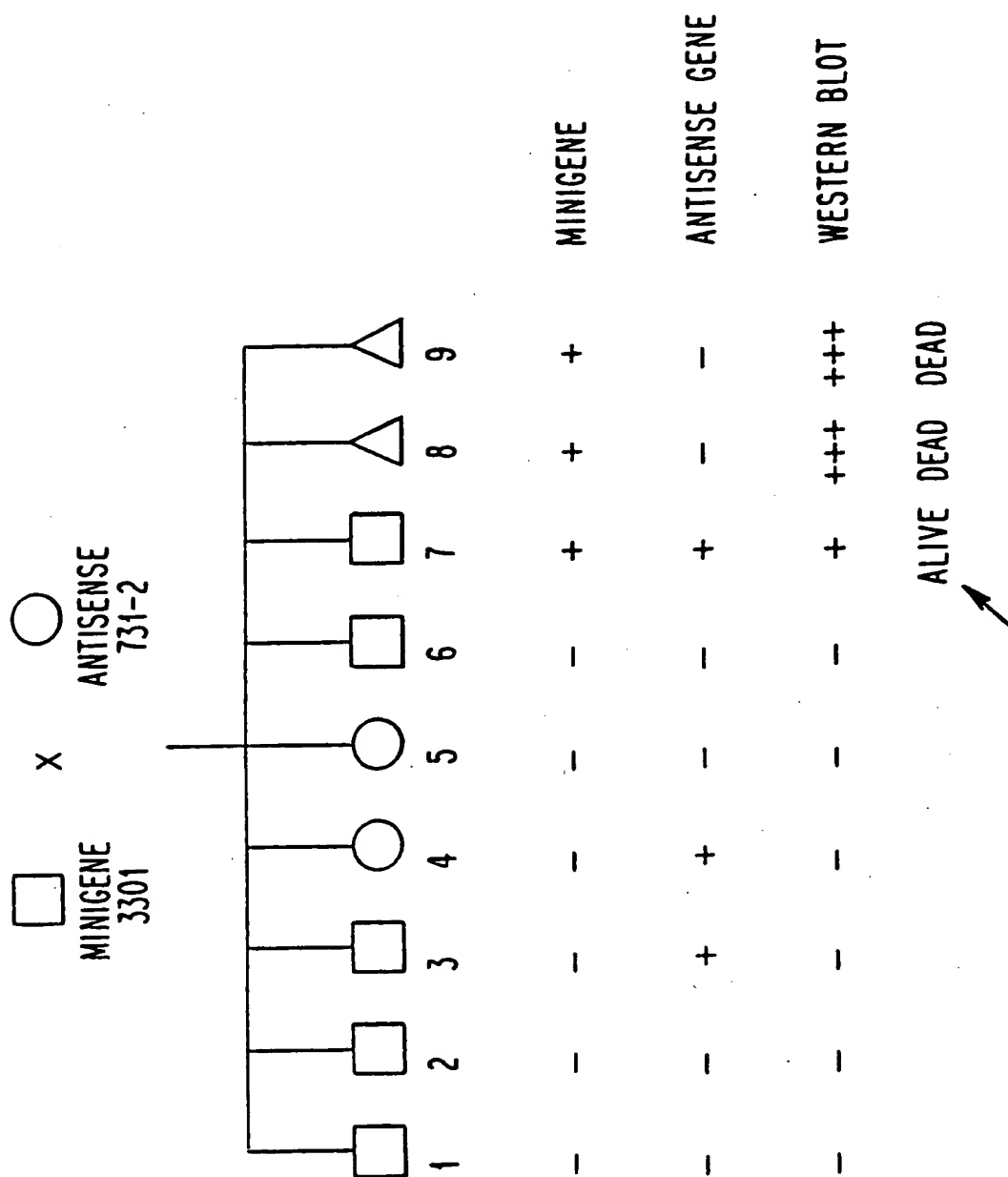
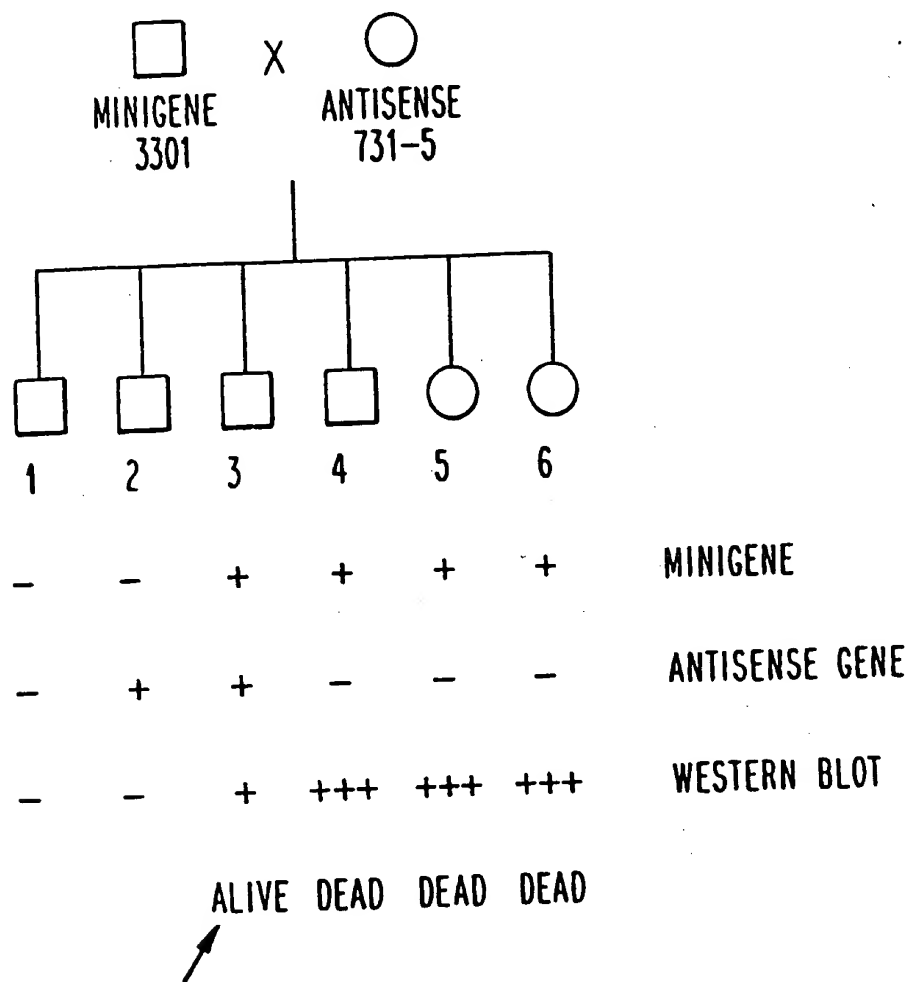


Fig. 1B

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*Fig. 2A*

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**Fig. 2B**

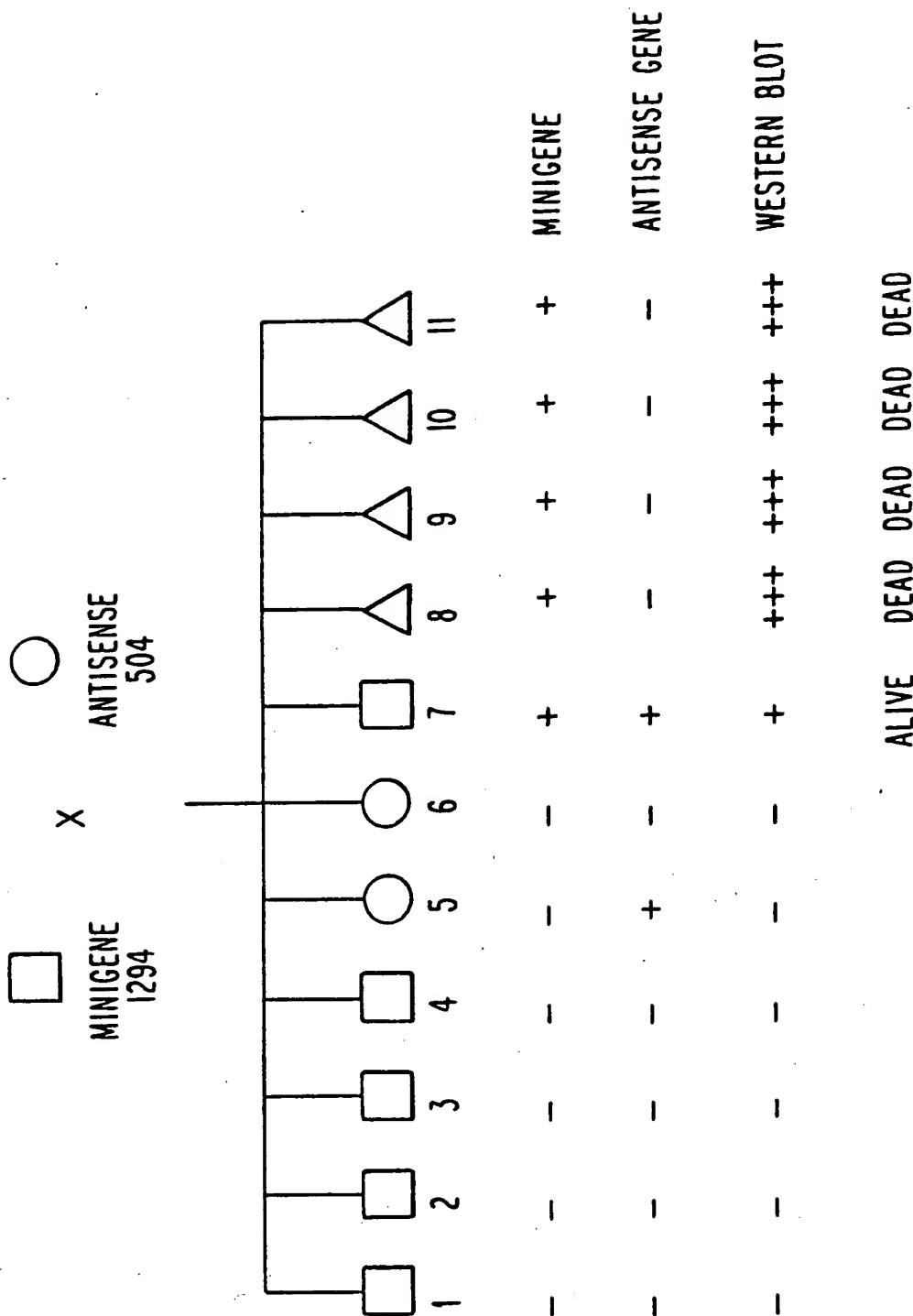


Fig. 2C

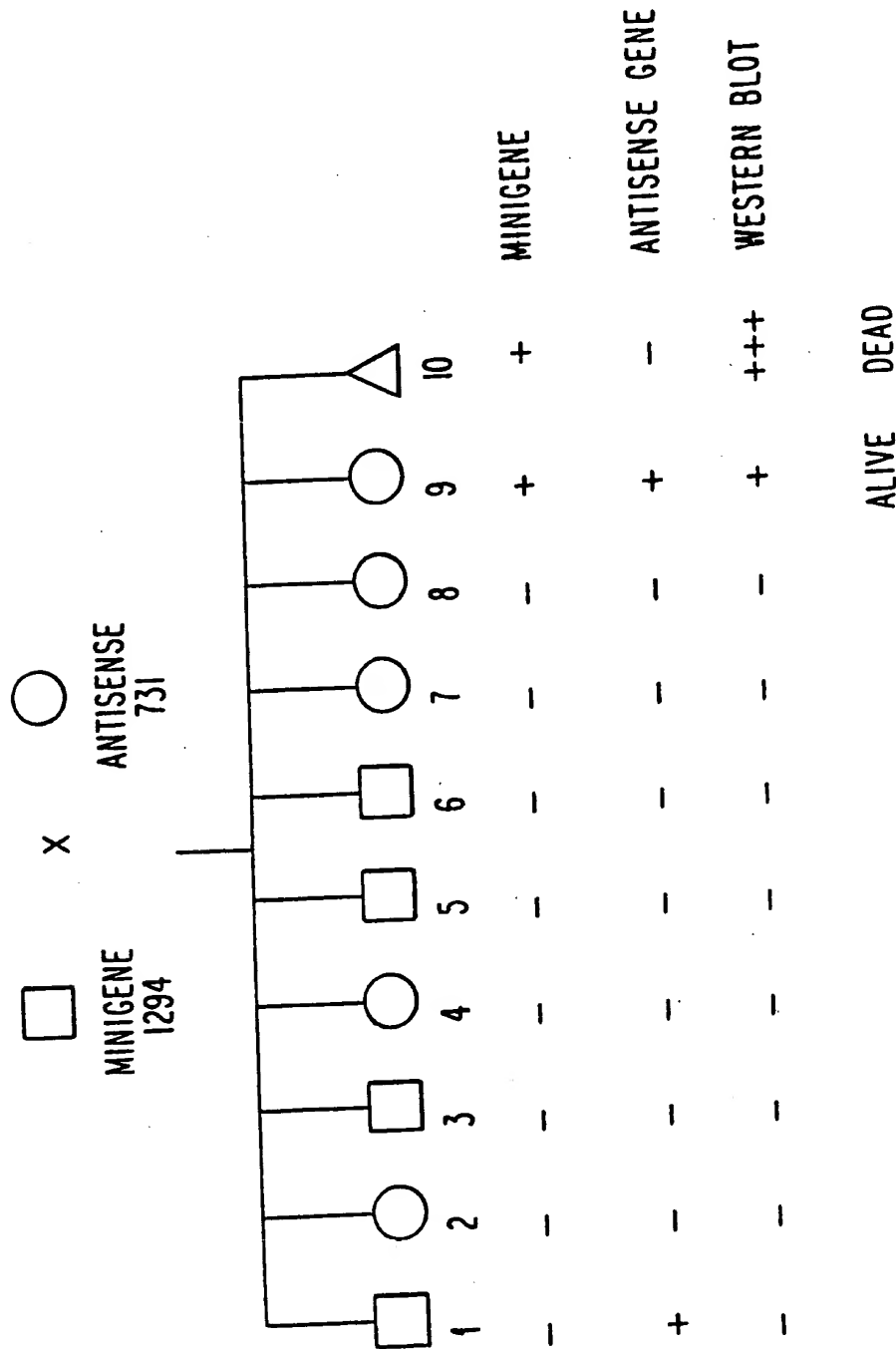


Fig. 2D

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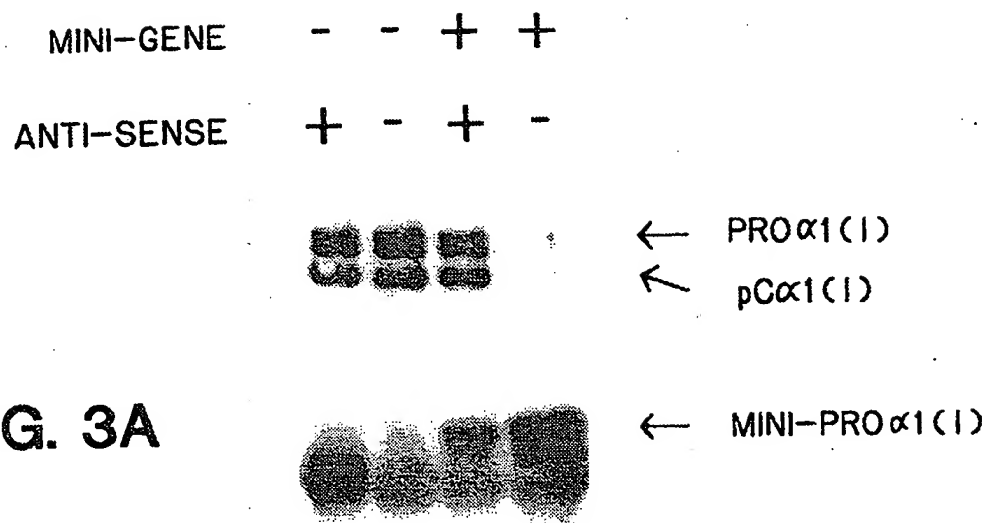


FIG. 3A

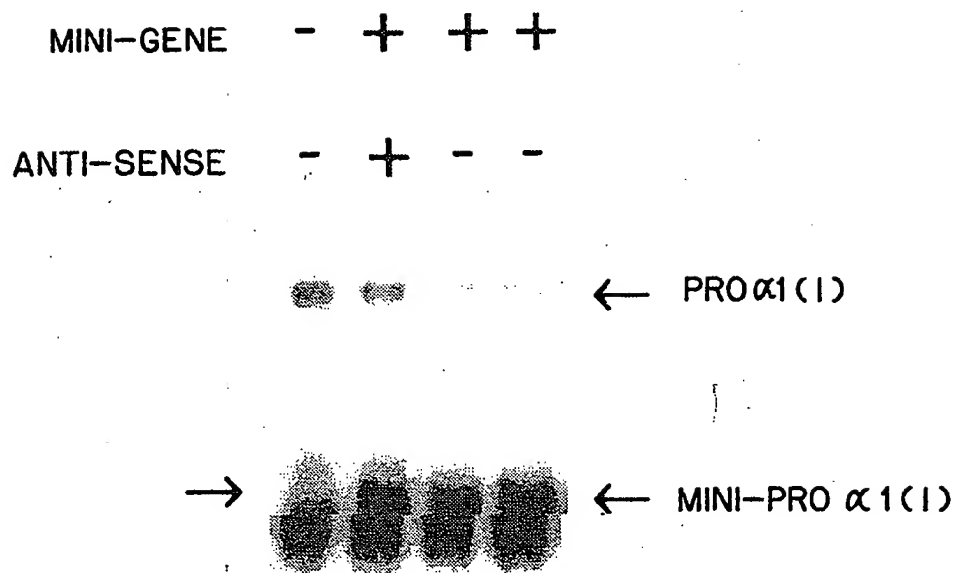


FIG. 3B

INTERNATIONAL SEARCH REPORT

International application No.
PCT/US94/03321

A. CLASSIFICATION OF SUBJECT MATTER

IPC(5) : A61K 48/00; C12N 5/16, 15/12, 15/63, 15/87, 15/88; C07H 21/04
US CL : 514/44; 424/93B; 435/240.2, 320.1, 172.3, 69.1; 536/23.5, 24.5

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

U.S. : 514/44; 424/93B; 435/240.2, 320.1, 172.3, 69.1; 536/23.5, 24.5

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)

BIOSIS, MEDLINE

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A	J. Biol. Chem., Vol. 266, No. 2, issued 15 January, 1991, Olsen et al., "High Levels of Expression of a Minigene Version of the Human Proalpha1(I) Collagen Gene in Stably Transfected Mouse Fibroblasts", pages 1117-1121, see entire document.	1-18
A	J. Biol. Chem., Vol. 266, No. 34, issued 05 December 1991, Khillan et al., "Transgenic Mice That Express a Minigene Version of the Human Gene for Type I Procollagen (COL1A1) Develop a Phenotype Resembling a Lethal Form of Osteogenesis Imperfecta", pages 23373-23379, see the entire document.	1-18

☒ Further documents are listed in the continuation of Box C.

☐ See patent family annex.

* Special categories of cited documents:

A document defining the general state of the art which is not considered to be of particular relevance

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O document referring to an oral disclosure, use, exhibition or other means

P document published prior to the international filing date but later than the priority date claimed

T

later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention

X

document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone

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document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art

G

document member of the same patent family

Date of the actual completion of the international search

20 JUNE 1994

Date of mailing of the international search report

JUL 05 1994

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INTERNATIONAL SEARCH REPORT

International application No.
PCT/US94/03321

C (Continuation). -DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A	The FASEB Journal, Vol. 5, issued April 1991, Kuivaniemi et al., "Mutations in collagen genes: causes of rare and some common diseases in humans", pages 2052-2060, see the entire document.	1-18
A	TIBTECH, Vol. 10, issued August 1992, Wickstrom, "Strategies for administering targeted therapeutic oligodeoxynucleotides", pages 281-287, see the entire document.	1-18